

Extraction of THC, THCA and Carboxy-THC from Oral Fluid by ISOLUTE® SLE+ after Collection with the Intercept® Oral Fluid Drug Test Kit Prior to GC/MS Analysis

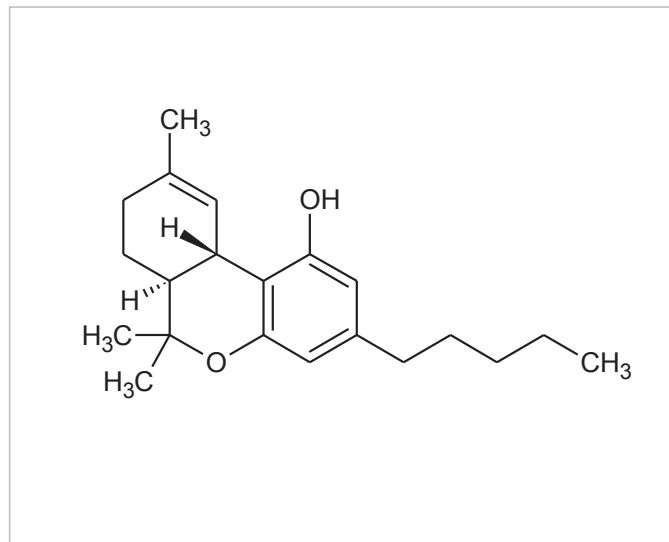


Figure 1. Structures of Δ^9 -THC (tetrahydrocannabinol)

Introduction

This application note describes the extraction of THC, THCA and Carboxy-THC from oral fluid matrix collected using the Intercept Oral Fluid Drug Test Kit (Orasure Technologies), prior to GC/MS analysis.

ISOLUTE® SLE+ Supported Liquid Extraction plates and columns offer an efficient alternative to traditional liquid-liquid extraction (LLE) for bio-analytical sample preparation, providing high analyte recoveries, no emulsion formation, and significantly reduced sample preparation.

This application note describes an effective and efficient ISOLUTE SLE+ protocol optimized for 400 μ L and 1 mL sample capacity formats. The simple sample preparation procedure delivers clean extracts and analyte recoveries greater than 80% with RSDs lower than 10% for all analytes.

Analytes

THC, THCA, THC-COOH and THC-d₃, THC-COOH-d₃ as internal standards

Sample Preparation Procedure

- Sample pre-treatment:** Following collection, add 0.5% ammonium hydroxide (aq) (10 μ L) to each collection device (see additional information).
- Format:** **ISOLUTE® SLE+ 400 μ L Supported Liquid Extraction columns, part number 820-0055-B**
- Sample loading:** Load the pre-treated oral fluid (300 μ L) onto the column and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.
- Analyte Extraction:** Apply dichloromethane/isopropanol, (95/5, v/v, 1 mL) and allow to flow under gravity for 5 minutes. Apply a further aliquot of DCM/IPA, (95/5, v/v, 1 mL) and allow to flow for another 5 minutes under gravity. Apply vacuum or positive pressure (5–10 seconds) to complete elution.
- Format:** **ISOLUTE® SLE+ 1 mL Supported Liquid Extraction columns, part number 820-0140-C**
- Sample loading:** Load the complete contents of the pre-treated oral fluid device onto the column and apply a pulse of vacuum or positive pressure (3–5 seconds) to initiate flow. Allow the sample to absorb for 5 minutes.
- Analyte Extraction:** Apply dichloromethane/isopropanol, (95/5, v/v, 2.5 mL) and allow to flow under gravity for 5 minutes. Apply a further aliquot of DCM/IPA, (95/5, v/v, 2.5 mL) and allow to flow for another 5 minutes under gravity. Apply vacuum or positive pressure (5–10 seconds) to complete elution.
- Post Elution & Reconstitution:** Dry the extract in a stream of air or nitrogen using a SPE Dry (40 °C, 20 to 40 L/min) or TurboVap (1.0 bar at 40 °C for 40 mins).
- Upon dryness, reconstitute with 50 μ L ethyl acetate and 25 μ L MTBSTFA:TBDMCS (99:1, v/v) and vortex for 20 seconds. Transfer to a high recovery glass vial. Place in a heating block set to 70 °C, for 25 minutes. Remove vial from the block and allow cooling.

GC Conditions

Instrument:	Agilent 7890A with QuickSwap
Column:	Phenomenex Zebron ZB-Semivolatiles, 30 m x 0.25 mm ID x 0.25 µm
Carrier	Helium 1.2 mL/min (constant flow)
Inlet:	250 °C, Splitless, purge flow: 50 mL/min at 1.0 min
Injection:	2 µL Wash solvents: ethyl acetate
Oven:	Initial temperature 100 °C Ramp 100 °C/min to 280 °C, hold for 10.5 minutes Ramp 100 °C/min to 330 °C, hold for 0.5 minutes
Post run:	Backflush for 2.4 minutes (3 void volumes)
Transfer Line:	280 °C

MS Conditions

Instrument:	Agilent 5975C
Source:	230 °C
Quadrupole:	150 °C
MSD mode:	SIM

SIM Parameters

Table 1. Ions acquired in the Selected Ion Monitoring (SIM) mode

SIM Group	Analyte	Target (Quant) Ion	1 st Qual Ion	2 nd Qual Ion
1	THC-d3	374	431	348
1	THC	371	428	345
2	THCA	530	631	455
3	THC-COOH-d3	416	518	575
3	THC-COOH	413	515	572

Results

The optimized ISOLUTE SLE+ protocol demonstrated analyte recoveries ranging from 76-94% as shown in **Figure 2**. RSDs were below 10% for all analytes.

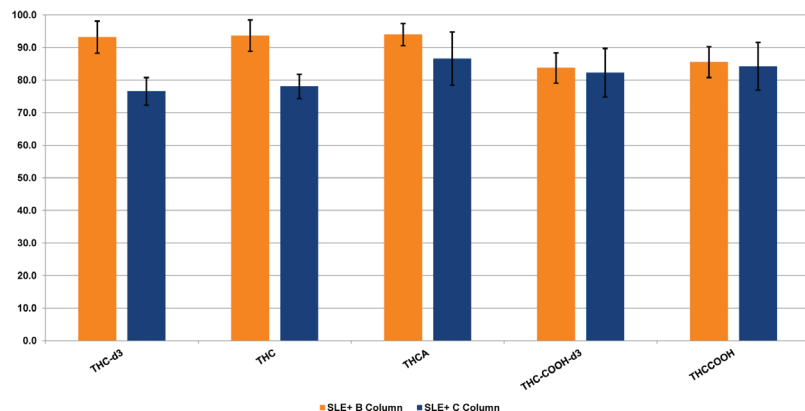


Figure 2. Typical extraction % recoveries (n=7) using the ISOLUTE® SLE+ protocol.

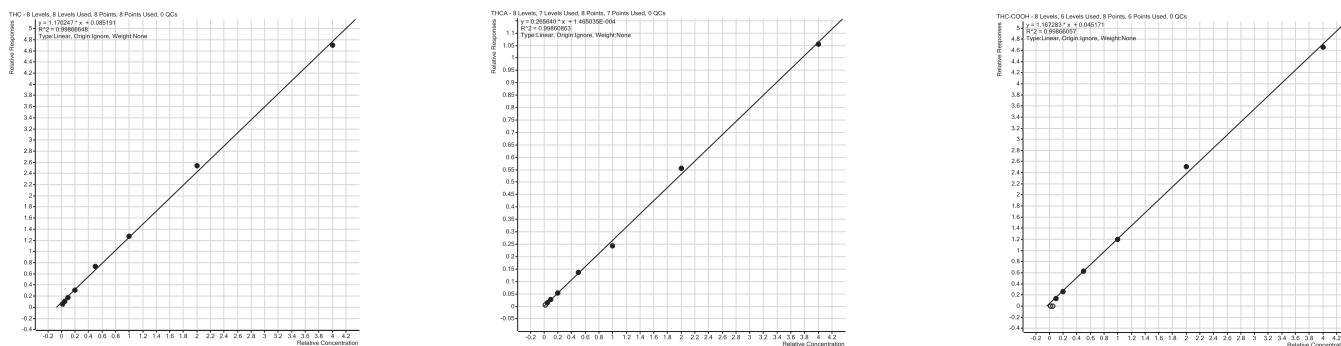


Figure 3. Calibration curves for extracted levels of spiked oral fluid after collection with Intercept devices, using 1 mL SLE+ format. Concentrations range from 4 ng/mL to 800 ng/mL showing r^2 values of approximately 0.9986 for the three analytes.

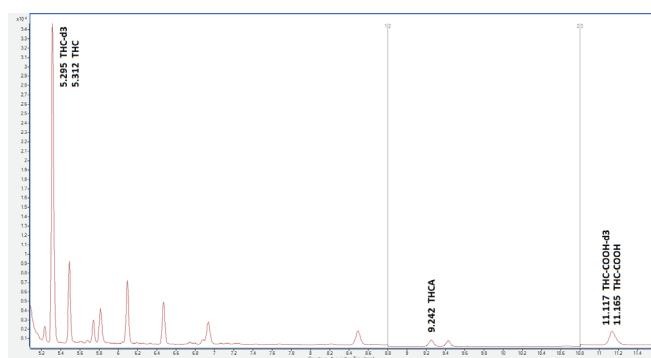


Figure 4. GC/MS chromatography for INTERCEPT collected oral fluid spiked at 40 ng/mL

Additional information

- 0.5 % ammonium hydroxide is prepared from concentrated stock (28-30%) by adding 50 μ L in 10 mL HPLC grade water.
- If a non-chlorinated solvent is required, MTBE (methyl-tert-butyl-ether) is a suitable alternative elution solvent.

Table 3. Lower Limits of Quantitation (LLOQ) using ISOLUTE® SLE+ procedure

Analyte	Lower Limit Of Quantitation
THC	4 ng/mL
THCA	10 ng/mL
THC-COOH	20 ng/mL

Ordering Information

Part Number	Description	Quantity
820-0055-B	ISOLUTE® SLE+ 400 µL Supported Liquid Extraction Columns	50
820-0140-C	ISOLUTE® SLE+ 1 mL Supported Liquid Extraction Columns	30
PPM-48	Biotage® PRESSURE+ 48 Positive Pressure Manifold 4	1
SD-9600-DHS-EU	Biotage® SPE Dry Sample Concentrator System 220/240 V	1
SD-9600-DHS-NA	Biotage® SPE Dry Sample Concentrator System 100/120 V	1
C103198	TurboVap® LV, 100/120V	1
C103199	TurboVap® LV, 220/240V	1

For the latest application notes visit www.biotage.com

EUROPE

Main Office: +46 18 565900
Toll Free: +800 18 565710
Fax: +46 18 591922
Order Tel: +46 18 565710
Order Fax: +46 18 565705
order@biotage.com
Support Tel: +46 18 56 59 11
Support Fax: +46 18 56 57 11
eu-1-pointsupport@biotage.com

NORTH & LATIN AMERICA

Main Office: +1 704 654 4900
Toll Free: +1 800 446 4752
Fax: +1 704 654 4917
Order Tel: +1 704 654 4900
Order Fax: +1 434 296 8217
ordermailbox@biotage.com
Support Tel: +1 800 446 4752
Outside US: +1 704 654 4900
us-1-pointsupport@biotage.com

JAPAN

Tel: +81 3 5627 3123
Fax: +81 3 5627 3121
jp_order@biotage.com
jp-1-pointsupport@biotage.com

CHINA

Tel: +86 21 2898 6655
Fax: +86 21 2898 6153
cn_order@biotage.com
cn-1-pointsupport@biotage.com

To locate a distributor,
please visit our website at
www.biotage.com

Part Number: AN819

© 2014 Biotage. All rights reserved. No material may be reproduced or published without the written permission of Biotage. Information in this document is subject to change without notice and does not represent any commitment from Biotage. E&OE. Product and company names mentioned herein may be trademarks or registered trademarks and/or service marks of their respective owners, and are used only for explanation and to the owners' benefit, without intent to infringe.

For more information visit www.biotage.com.